

Evaluation of LAMP-TB system for detecting *Mycobacterium tuberculosis*

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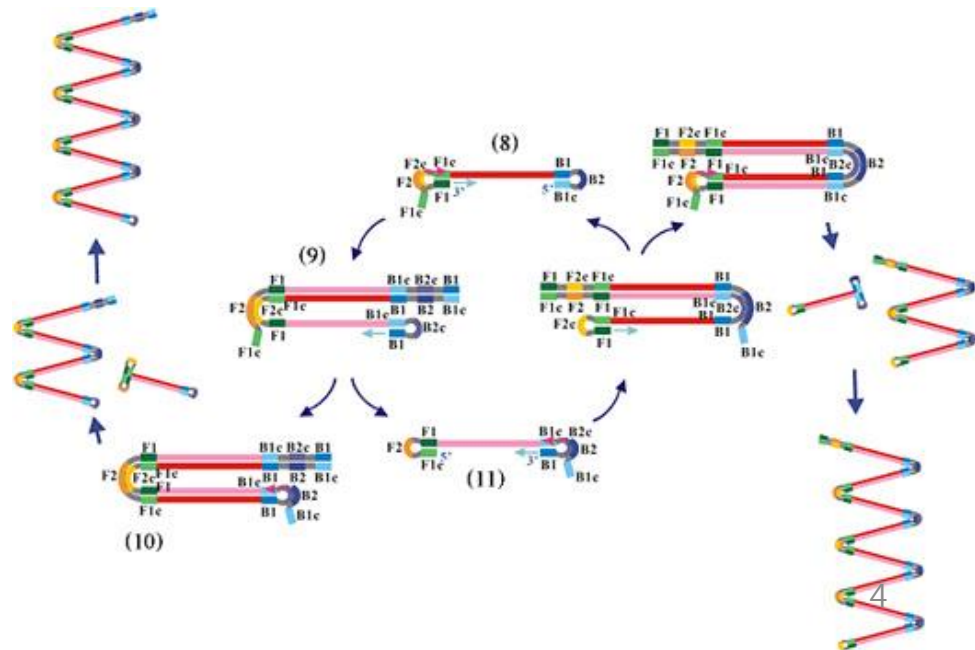
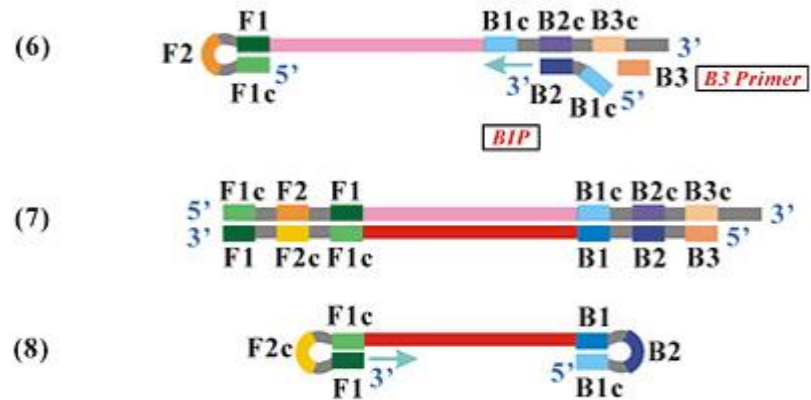
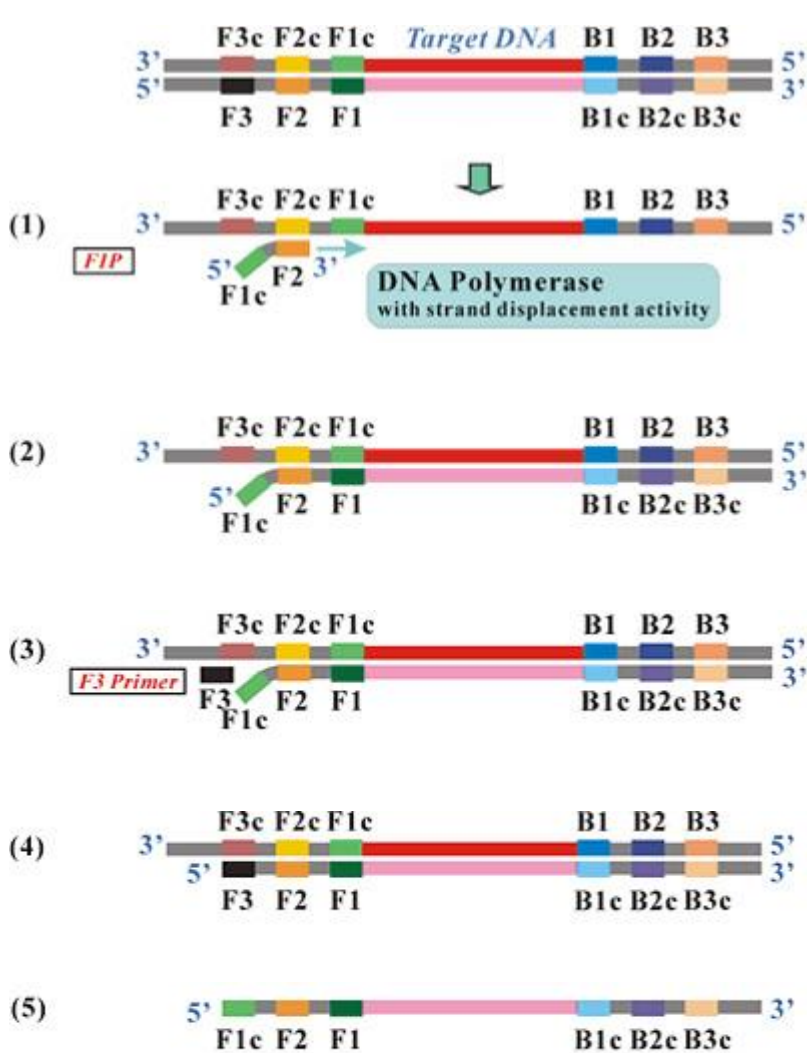
Introduction

- Laboratory tests are crucial for tuberculosis control.
 - Detection, treatment monitoring & drug susceptibility testing
- Limitations of conventional tests for TB
 - Smear microscope: low sensitivity & specificity
 - Culture: complex & long TAT
- Limitations of current molecular tests
 - Require facilities & equipment
 - Require technical expertise & human resources
 - Often not feasible in resource-poor settings or remote area

Introduction

- Loop-mediated isothermal amplification (LAMP)
 - Simple and rapid nucleic acid amplification method
 - Using 4 primers for 6 regions and one enzyme
 - Isothermal amplification
- LAMP-TB system
 - Detecting MTB using LAMP method in clinical sample
 - Reaction time: ~60 mins
 - Simple DNA extraction kit / Premix reaction tube
 - Mitarai et al.: sensitivity - 98.2% for Sm(+)/Cx(+), 55.6% for Sm(-)/Cx(+)

Introduction-LAMP



Global TB diagnostic pipeline

| | Early Development | Late or Completed Development | On Pathway to WHO Evaluation |
|---|---|--|--|
| HIGH COMPLEXITY ASSAYS | Molecular Detection/DST | | |
| | TruArray MDR-TB (Akkoni) COBAS TaqMan MTB +DST(Roche) Hydra 1K (insilixa) Mycobacterium Real-time MDR (CapitalBio) | TRC Rapid MTB (Tosoh) VereMTB (Veredus Laboratories) LiPA Pyrazinamide (Nipro) LATE-PCR Lights on / Lights off (Hain) TBMDx (Abbott) Meltpro (Zeesan) Mycobacteria RT PCR (CapitalBio) REBA MTB-XDR (YD Diagnostics) EasyNAT TB (Ustar) BD Max (BD) | GenoTYPE MTBDRsl (Hain) LiPA MDR-TB (Nipro) REBA MTB-Rifa (YD Diagnostics) |
| | Culture-based Technology | | |
| | BNP Middlebrook (NanoLogix) Rapid colorimetric DST | TREK Sensitive MYCOTB (Trek) | |
| MODERATE COMPLEXITY ASSAYS | Molecular Detection/DST | | |
| | Xpert Ultra and Xtend XDR (Cepheid) Alere Q (Alere) Enigma ML (Enigma Diagnostics) Q-POC (QuantuMDx) EOSCAPE (Wave80) RT-PCR Testing Platform (NWGHF/Guidel) iCubate 2.0 (iCubate) TBDx system (KGI) DiagCORE (STAT Diagnostica) LabChip G2-3 (Nanobiosys) | Genedrive MTB/RIF (Epistem) Truelab/Truenat MTB (Molbio) | TB LAMP (Eiken) |
| | Volatile Organic Compounds | | |
| | BreathLink (Menssana) Prototype breathalyzer (Next Dimensions) TB Breathalyser (Rapid Biosensor Systems) Aeonose (The eNose Company) Breath analysis instrument (Metabolomx) | Giant African Pouch Rats (Apopo) | |
| Automated Microscopy & Imaging | | | |
| TBDx (Applied Visual Sciences) Fluorescent microscopy (ID-FISH Tech.) Automatic TB Screener (Fluorobot) | Microimager (BD) CAD4TB (Delft Imaging Systems) | | |



Purpose of the study

- Evaluate the performance of LAMP-TB system for detecting TB comparing with conventional tests and real-time PCR

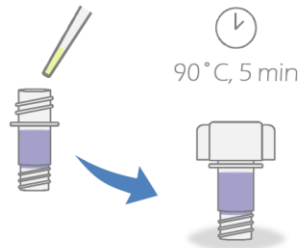
Materials and Methods

- **Specimen:** 226 sputum samples from TB suspects of health centers
- **Conventional tests**
 - Smear microscope: auramine O with WHO grading
 - Culture: solid culture (2% Ogawa), liquid culture (MGIT 960)
- **Real-time PCR**
 - DNA extraction: heat-extraction
 - Reagent: Advansure MTB/NTM kit (LG lifescience, Korea)
- **LAMP-TB**
 - DNA extraction: Loopamp™ Pure DNA extraction kit
 - Reagent: Loopamp™ MTBC detection kit
 - Amplification and detection: LA-500 (Eiken, Japan)

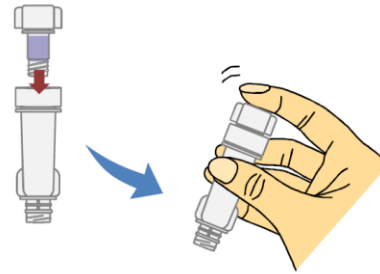
DNA Extraction

(Loopamp™ PURE DNA Extraction Kit)

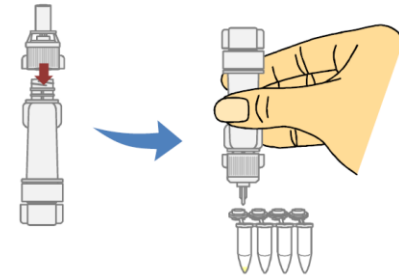
- 1 Pipette sputum into Heating Tube and incubate at 90°C for 5 minutes



- 2 Connect Heating Tube to Adsorbent Tube and mix completely by shaking



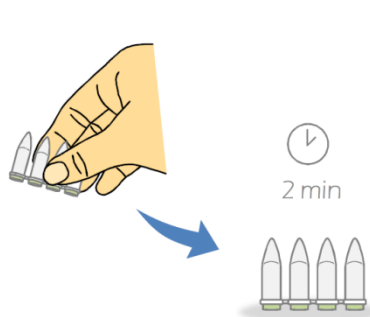
- 3 Attach Injection Cap. Squeeze and drop sample into reaction tube



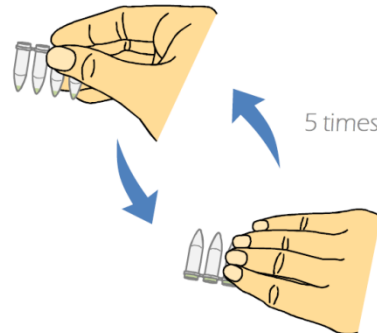
Amplification and Detection

(Loopamp™ MTBC Detection Kit)

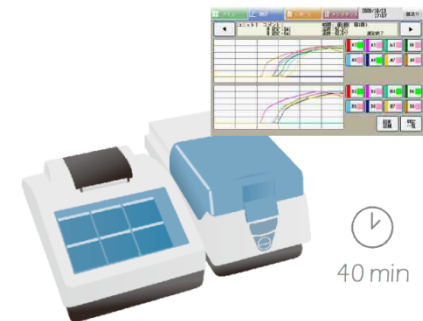
- 1 Invert reaction tubes to collect solution in cap. Leave tubes standing upside down for 2 minutes



- 2 Mix solution with inversion of tubes and spin down tubes



- 3 Set reaction tubes to turbidimeter or incubator and start LAMP reaction



Culture vs LAMP-TB

| Culture | LAMP-TB | | Total |
|---------------|--------------------|-------------------|------------|
| | Negative | Positive | |
| MTB | 4 (26.7%) | 11 (73.3%) | 15 |
| NTM | 33 (100%) | - | 33 |
| No growth | 154 (97.5%) | 4 (2.5%) | 158 |
| Contamination | 1 (100%) | - | 1 |
| Pending | 2 (10.5%) | 17 (89.5%) | 19 |
| Total | 194 (85.8%) | 32 (14.2%) | 226 |

Sm(+)/Cx(+) = 87.5% (7/8), Sm(-)/Cx(+) = 57.1% (4/7)

LAMP-TB results according to Smear grade

| Smear results | Positivity of LAMP-TB | Total |
|----------------|-----------------------|-------|
| Negative | 5 (3.2%) | 157 |
| 1+ | 18 (72.0%) | 25 |
| 2+ | 5 (71.4%) | 7 |
| 3+ | 4 (100%) | 4 |
| Total | 32 (16.6%) | 193 |
| Smear positive | 27 (75.0%) | 36 |

※ 33 samples which were culture positive for NTM were not included.

LAMP-TB vs Real-time PCR

| Real-time PCR | LAMP-TB | | Total |
|---------------|--------------------|-------------------|------------|
| | negative | Positive | |
| MTB | 6 (15.8%) | 32 (84.2%) | 38 |
| MIX | 1 (100%) | - | 1 |
| NTM | 33 (100%) | - | 33 |
| Negative | 154 (100%) | - | 154 |
| Total | 194 (85.8%) | 32 (14.2%) | 226 |

Correlation between real-time PCR ct value & LAMP-TB

| ct value | LAMP-TB | | Total |
|----------|-----------|----------|-------|
| | Positive | Negative | |
| <30 | 30 (96.8) | 1 (3.2) | 31 |
| ≥30 | 2 (25.0) | 6 (75.0) | 8 |
| Total | 32 (82.1) | 7 (17.9) | 39 |

Conclusions

- LAMP-TB system is easy to perform with very low sample volume.
- Sensitivity and specificity of LAMP were similar with previous studies.
- Sensitivity of LAMP is less than that of real-time PCR.
- No. of samples positive for MTB was not enough for evaluation. The further study is needed.